

# AMPLIRUN® CHLAMYDIA TRACHOMATIS DNA CONTROL

For *in vitro* diagnostic use

**MBC012:** Purified DNA of *Chlamydia trachomatis* to be used to control *in vitro* diagnosis techniques based in nucleic acids amplification.

## INTRODUCTION:

Chlamydiae are nonmotile, obligate intracellular bacteria with a unique life cycle that includes two phases: reticulate and elementary bodies. *Chlamydia trachomatis* is comprised of two human biovars: the lymphogranuloma venereum, remarkable for its tropism for lymphoid cells and its ability to cause systemic disease, and the trachoma biovar, limited primarily to epithelial cells of mucous membranes and able to cause trachoma, sexually transmitted disease, and neonatal inclusion conjunctivitis and pneumonia.

## CHARACTERISTICS:

The lyophilized nucleic acid is included in a thermo-sealed foil pouch containing a silica gel bag. It is necessary to reconstitute it before use (refer to "Preparation of reagents").

**Preparation:** Purified from McCoy infected cells by sonication and differential centrifugation

**Extract preparation:** Commercial genomic DNA extraction method.

## KIT CONTENTS:

1 VIRCELL CTR DNA CONTROL: 1 vial with lyophilized DNA of *Chlamydia trachomatis*, (Lymphogranuloma venereum (LGV II) 434 strain), (10000-20000 copies/μl once reconstituted (see Table 1)). DNA quantification has been performed by real-time PCR.

2 VIRCELL CONTROL RECONSTITUTION SOLUTION: 500 μl of molecular biology grade water, DNase, RNase free.

Lot number	
Concentration	copies/μl

Table 1.

Materials required but not supplied:

Additional diagnosis kit.

## STORAGE REQUIREMENTS:

Special transport conditions not required. Store the lyophilized vial at 2-8°C inside the foil pouch. Once the pouch is opened, reconstitute the lyophilized vial immediately and store between -5°C and -40°C after reconstitution (temperature indicated on the label).

## STABILITY AND HANDLING OF REAGENTS:

Handle reagents in aseptic conditions to avoid microbial contaminations.

Use only the amount of reagent required for the test.

After control resuspension DNA solution should be aliquoted in order to avoid multiple freeze-thaw cycles. The product is stable until the expiry date indicated in the label, if the instructions for use are followed.

VIRCELL, S.L. does not accept responsibility for the mishandling of the reagents included in the kit.

## RECOMMENDATIONS AND PRECAUTIONS:

1. This product is for *in vitro* diagnosis use only and for professional qualified staff.
2. Sterile tips with aerosol barrier are essential to prevent contamination.
3. Specimens should be handled as in the case of infectious samples using safety laboratory procedures. Thoroughly clean and disinfect all work surfaces with a freshly prepared solution of 0.5% sodium hypochlorite in deionized or distilled water.
4. In order to perform the test it is essential to have separate working areas.
5. Dispose of unused reagents and waste in accordance with all applicable regulations.
6. The component VIRCELL DNA CONTROL could include genetic material or substances of animal and/or human origin. VIRCELL DNA CONTROL could contain inactivated *Chlamydia trachomatis* antigen. VIRCELL DNA CONTROL contains purified nucleic acids obtained from inactivated microorganism, nevertheless, it should be considered potentially infectious and handled with care. No present method can offer complete assurance that these or other infectious agents are absent. All materials should be handled and disposed as of potentially infectious. Observe the local regulations for clinical waste disposal.
7. Dilutions below 1000 copies/μl should be made immediately before use. Freezing of product dilutions containing less than 1000 copies/μl is not recommended as partial DNA degradation might occur.

## PREPARATION OF THE REAGENTS:

1. Tear the foil pouch containing VIRCELL DNA CONTROL 1.
2. Centrifuge VIRCELL DNA CONTROL 1 1 minute at 1000 g.
3. Add 100 μl of VIRCELL CONTROL RECONSTITUTION SOLUTION 2 and mix until completely reconstituted. The concentration will be 10000-20000 copies/μl once reconstituted (see Table 1).
4. Shake with vortex for 30 seconds to dissolve and homogenize completely.
5. It is recommended to prepare VIRCELL DNA CONTROL aliquots. In case dilutions were to be prepared use VIRCELL CONTROL RECONSTITUTION SOLUTION 2 for this purpose.

## TEST PROCEDURE:

Once nucleic acid is reconstituted, use it according to indications of additional diagnosis kit. Use resuspended VIRCELL CONTROL as an extracted clinical sample adding it directly to amplification reagents.

## INTERNAL QUALITY CONTROL:

Each batch is subjected to internal quality control testing before releasing. Quality control analysis is performed by real-time PCR. Final quality control results for each particular lot are available.

## INTERPRETATION OF RESULTS AND VALIDATION PROTOCOL FOR USERS:

Refer to indications of additional diagnosis kit.



**LIMITATIONS OF METHOD:**

1. This reagent is intended to be used with methods of human diagnosis. This test has not been verified with other methods.
2. The user of this kit is advised to read carefully and understand the package insert. Strict adherence to the protocol is necessary to obtain reliable test results.
3. Use of this product should be limited only to personnel trained in molecular techniques.
4. The identity test was carried out with some specific primers according to publicly available sequences of the microorganism. Changes in the sequences of the primers of the reaction may produce a range of different sizes or may not display product amplification.
5. This control does not substitute internal diagnostic kit controls.
6. The quantification was carried out by own brand qPCR against a standard used as a calibrator. Results may vary with the amplification conditions of the end user.
7. AMPLIRUN® has not been designed to be used with a particular diagnostic kit coming from a certain manufacturer. It is used to control amplification of a diagnostic laboratory functioning procedure.

**PERFORMANCES:****• IDENTITY TEST**

**PCR analysis of DNA control:** PCR analysis was performed with a specific oligonucleotide pair on purified *Chlamydia trachomatis* DNA. The reaction produced a fragment of the expected size.

**• QUANTIFICATION TEST**

A correlation test was performed between microorganism culture and *Chlamydia trachomatis* extracted DNA. Less than 0.5 log variance was observed between both assays.

**• INTRA-ASSAY PRECISION**

3 replicas of 5 serial dilutions of 3 different vials of the product were performed by the same operator under identical qPCR conditions.











Less than 5% coefficient of variance was observed between all assays.

**• INTER-ASSAY PRECISION**

3 different replicas of 5 different serial dilutions of 1 vial of the product were individually amplified by 2 different operators on 3 consecutive days.

Less than 5% coefficient of variance was observed between all assays.

**SYMBOLS USED IN LABELS:**

	In vitro diagnostic medical device
	Use by (expiration date)
	Store at x-y°C
	Batch code
	Catalogue number
	Consult instructions for use
	Reconstitute in x µl
	Shipment temperature
	Storage temperature
	Do not open until use

**BIBLIOGRAPHY:**

1. Bustin SA. (2000). Absolute quantification of mRNA using real-time reverse transcription polymerase chain reaction assays. J Mol Endocrinol. 25(2):169-193.
2. Bustin SA. (2002). Quantification of mRNA using real-time reverse transcription PCR (RT-PCR): trends and problems. J Mol Endocrinol. 29(1):23-39.
3. Freeman WM, Walker SJ, Vrana KE. (1999). Quantitative RT-PCR: pitfalls and potential. Biotechniques. 1999 26(1):112-122, 124-125.
4. Jatón K., Bille J., and Greub G. (2006). A novel real-time PCR to detect *Chlamydia trachomatis* in first-void urine or genital swabs. Journal of Medical Microbiology, 55,1667-1674.
5. Larionov A, Krause A, Miller W. (2005). A standard curve based method for relative real time PCR data processing. BMC Bioinformatics. 6:62.
6. Madico G, Quinn TC, Boman J, Gaydos CA. (2000). Touchdown enzyme time release-PCR for detection and identification of *Chlamydia trachomatis*, *C. pneumoniae*, and *C. psittaci* using the 16S and 16S-23S spacer rRNA genes. J Clin Microbiol. 2000 38(3):1085-10.

For further inquiries please contact:

[customerservice@vircell.com](mailto:customerservice@vircell.com)

REVISED: 2019-01-15

L-MBC012-EN-01

